



### **Test Characteristics**

Test Name Potato mop-top virus Test Label FAM-labeled target probe

Catalog Number 12501 Internal Control Endogenous

Acronym PMTV Format XRT

Genus Pomovirus Diluents GEB/PD1

Binomial Name Pomovirus solani Sample Dilution 1:10

### Summary

AmplifyRP XRT for PMTV is a rapid RNA amplification and detection platform designed for testing potato leaves, stems, tubers, or tissue culture plantlets for Potato mop-top virus. This kit includes lyophilized reaction pellets containing the necessary reagents to amplify PMTV RNA and an endogenous RNA control at a single operating temperature (42 °C).

### Diagnostic Sensitivity Analytical Sensitivity

True Positives 30 Analytical Sensitivity: The assay is 87.5% sensitive between 1 fg/ $\mu$ L and 10 fg/ $\mu$ L. (n=16)

Correct Diagnoses 30 Limit of Detection: The assay has a 100% detection rate at 10 fg/µL with RNA transcripts. (n=8)

**Percent** 100% The assay has a 75% detection rate at 1 fg/ $\mu$ L with RNA transcripts. (n=8)

### **Analytical Specificity**

### Inclusivity:

### Isolates and Geographic Regions Detected:

TV-23T (Colombia) <sup>1</sup> PMTV-C52_P11 (Peru) <sup>1</sup>		
PMTV-Guangdong (China) <sup>1</sup>	PMTV-ID-Idaho_1 (USA)¹	
PMTV-J20_P157 (Peru) <sup>1</sup>	PMTV-J21_P117 (Peru) <sup>1</sup>	
PMTV-Maryland (USA) <sup>1</sup>	PMTV-Nd4 (Canada) <sup>1</sup>	
PMTV-physalis_M7 (Colombia) <sup>1</sup>	PMTV-PV-0582 (UK)	
PMTV-quitoense_M6 (Colombia)¹ PMTV-Swedish, (Sw) (Sweden)¹		
PMTV-Todd <sup>1</sup>	PMTV-Yunnan (China) <sup>1</sup>	
<sup>1</sup> Predicted detection by in silico analysis only		

# Exclusivity:

#### Cross-reacts With:

Virus Name	Species Name
None Known	

#### Does Not Cross-react With:

Virus Name	Species Name
Alfalfa mosaic virus (AMV)	Alfamovirus AMV
Andean potato latent virus (APLV) <sup>1</sup>	Tymovirus latandigenum
Andean potato mottle virus (APMoV)	Comovirus andesense

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### Does Not Cross-react With:

Does Not Cross-react With:		
Virus Name	Species Name	
Barley stripe mosaic virus (BSMV) <sup>1</sup>	Hordeivirus hordei	
Beet soil-borne virus (BSBV)	Pomovirus solibetae	
Beet virus Q (BVQ) <sup>1</sup>	Pomovirus betae	
Broad bean necrosis virus (BBNV) <sup>1</sup>	Pomovirus viciae	
Colombian potato soil-borne virus (CPSbV) <sup>1</sup>	Pomovirus colombiense	
Cucumber green mottle mosaic virus (CGMMV) <sup>1</sup>	Tobamovirus viridimaculae	
Indian peanut clump virus (IPCV) <sup>1</sup>	Pecluvirus indicum	
Japanese soil-borne wheat mosaic virus (JSBWMV) <sup>1</sup>	Furovirus japonicum	
Pea early-browning virus (PEBV) <sup>1</sup>	Tobravirus pisi	
Peanut clump virus (PCV) <sup>1</sup>	Pecluvirus arachidis	
Pepper ringspot virus (PepRSV) <sup>1</sup>	Tobravirus capsici	
Potato aucuba mosaic virus (PAMV)	Potexvirus marmoraucuba	
Potato latent virus (PotLV)	Carlavirus latensolani	
Potato leafroll virus (PLRV)	Polerovirus PLRV	
Potato virus A (PVA)	Potyvirus atuberosi	
Potato virus M (PVM)	Carlavirus misolani	
Potato virus S (PVS)	Carlavirus sigmasolani	
Potato virus T (PVT)	Tepovirus tafsolani	
Potato virus V (PVV)	Potyvirus vetuberosi	
Potato virus X (PVX)	Potexvirus ecspotati	
Potato virus Y (PVY)	Potyvirus yituberosi	
Rehmannia mosaic virus (ReMV)¹	Tobamovirus rehmanniae	
Ribgrass mosaic virus (RMV) <sup>1</sup>	Tobamovirus plantagonis	
Soil-borne wheat mosaic virus (SBWMV) <sup>1</sup>	Furovirus tritici	
Tobacco etch virus (TEV)	Potyvirus nicotianainsculpentis	
Tobacco mild green mosaic virus (TMGMV) <sup>1</sup>	Tobamovirus mititessellati	
Tobacco mosaic virus (TMV)¹	Tobamovirus tabaci	
Tobacco rattle virus (TRV)	Tobravirus tabaci	
Tomato brown rugose fruit virus (ToBRFV) <sup>1</sup>	Tobamovirus fructirugosum	
Tomato mosaic virus (ToMV)¹	Tobamovirus tomatotessellati	
Tomato spotted wilt virus (TSWV)	Orthotospovirus tomatomaculae	
¹Predicted non-detection by in silico analysis only		

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### **Diagnostic Specificity**

True Negatives 66
Correct Diagnoses 66
Percent 100%

### Selectivity:

No Matrix Effect Observed With:			
Potato leaves	Potato sprouts	Potato tubers	

The hosts on the above list have been chosen to represent those which historically cause a range of matrix effects, in addition to those expected to be screened for this pathogen. Not all plant species susceptible to this pathogen have been screened, but may still be used with this assay unless otherwise noted below. As with all diagnostic tools, Agdia recommends confirming all results with a secondary detection method before making any economic decisions (ex: discarding plants due to positive test results, etc.).

Matrix Effect Observed With:			
None Known			

# Repeatability

# Reproducibility

Number of Samples	88	Number of Samples	33
Replicates per Sample	2 - 3	Replicates per Sample	2 - 3
Total Replicates	190	Number of Operators	3
Replicates in Agreement	187	Total Replicates	198
Percent Agreement	98.4%	Replicates in Agreement	182
		Percent Agreement	91.9%

### **Robustness**

#### Planned deviation analysis:

No deviations from the user guide protocol were validated.

### Stability:

	1-year stability (accelerated)	Real-time Stability Verification
Positive Sample (High)	Pass	Monitoring
Positive Sample (High)	Pass	Monitoring
Positive Sample (Low)	Pass	Monitoring
Positive Sample (Low)	Pass	Monitoring
Positive Sample (Low)	Pass	Monitoring
Positive Sample (Low)	Pass	Monitoring
Negative Sample	Pass	Monitoring
Negative Sample	Pass	Monitoring

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#### Glossary

Diagnostic sensitivity': The percentage of positive samples correctly identified in an experiment with known positive controls.

Diagnostic specificity': The percentage of negative samples correctly identified in an experiment with known negative controls.

Analytical sensitivity3: The smallest amount of target that can be detected reliably (this is sometimes referred to as the 'limit of detection')

Analytical specificity<sup>3</sup>: (comprises inclusivity and exclusivity)

Inclusivity<sup>3</sup>: The performance of a test with a range of target isolates covering genetic diversity, different geographical origin and/or hosts

associated with the target organism.

Exclusivity<sup>3</sup>: The performance of a test with a range of non-targets (e.g. cross-reaction with closely related organisms, contaminants)

Selectivity<sup>2</sup>: The level of effect that matrices and relevant plant parts have on the performance of the assay.

Repeatability<sup>2</sup>: The agreement between test replicates of the same sample tested by the same operator.

Reproducibility<sup>3</sup>: The ability of a test to provide consistent results when applied to aliquots of the same sample tested under different conditions

(e.g. time, users, equipment, location)

Robustness<sup>1,3</sup>: The extent to which varying test conditions (e.g. temperature, volume, change of buffers) affect the established test performance

values. May also be referred to as planned deviation analysis.

Stability<sup>1</sup>: The performance of test reagents or controls over time.

References:

<sup>1</sup>Groth-Helms, D., Rivera, Y., Martin, F. N., Arif, M., Sharma, P., Castlebury, L. A. (in press). Terminology and Guidelines for Diagnostic Assay Development and Validation: Best Practices for Molecular Tests. PhytoFrontiers.

<sup>2</sup>Eads, A., Groth-Helms, D., Davenport, B., Cha, X., Li, R., Walsh, C., Schuetz, K., (in press). The Commercial Validation of Three Tomato Brown Rugose Fruit Virus Assays. PhytoFrontiers.

<sup>3</sup>EPPO (2018) PM 7/76 (5) Use of EPPO Diagnostic Standards, EPPO Bulletin 48, 373–377.

#### **Questions or Technical Support:**

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AmplifyRP Test Kits employ recombinase polymerase amplification (RPA) technology, developed by TwistDx Limited, U.K. Use of the RPA process and probe technologies are protected by US patents 7,270,981 B2, 7,399,590 B2, 7,435,561 B2, 7,485,428 B2 and foreign equivalents in addition to pending patents.

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